

# **SHORT PROTOCOL MAQ Control**

## **Preparation of Master Reaction Mix**

- 1. Remove the PCR mix from the -20 °C freezer and allow complete thawing on ice.
- 2. Vortex thoroughly and centrifuge the tubes at  $12,000 \times g$  for  $10 \times g$  before use.
- 3. Mix together **3 μl** PCR mix with **2 μl** optimized primer mix (for optimization, see user guide You MAQ) and **0.075 μl** Tag DNA polymerase.
- 4. Vortex briefly and centrifuge the tubes at  $12,000 \times g$  for  $10 \times s$ .

#### **PCR Reaction Setup**

- 5. For each sample, combine **20-50 ng** genomic DNA with **5 \muI** of the master reaction mix.
- 6. Adjust with distilled water to final volume of  $15 \mu l$ .
- 7. Vortex briefly and centrifuge the tubes at 12,000 x g for 10 s.

## **PCR Cycling Profile**

Note: Keep the ramp rate of the machine below 2.5 °C/s.

# **Fragment Analysis**

- 8. Prepare the size standard mix by combining **10 μl** of HiDi-Formamide (not provided) with **0.3 μl** GS500-liz size standard (not provided).
- 9. For each reaction, dispense **10**  $\mu$ **I** of size standard mix per well into a 96-well plate.
- 10. Add  $\mathbf{2} \mu \mathbf{I}$  of the MAQ PCR product to a well containing the size standard mix.
- 11. Denature samples at 95 °C for 3 min and put on ice immediately.
- 12. Centrifuge the plate at  $1,000 \times g$  for  $10 \times s$ .
- 13. Load onto the fragment analyzer.
- Analyse data with MAQ-S analysis software (freely downloadable from www.agilent.com)

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